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Research

Circum-Arctic distribution of chemical anti-herbivore compounds suggests biome-wide trade-off in defence strategies in Arctic shrubs

Elin Lindén, Mariska te Beest, Ilka Aubreu, Thomas Moritz, Maja K. Sundqvist, Isabel C. Barrio, Julia Boike, John P. Bryant, Kari Anne Bråthen, Agata Buchwal, C. Guillermo Bueno, Alain Currier, Dagmar D. Egelkraut, Bruce C. Forbes, Martin Hallinger, Monique Heijmans, Luise Hermanutz, David S. Hik, Annika Hofgaard, Milena Holmgren, Diane C. Huebner, Toke T. Høye, Ingibjörg S. Jónsdóttir, Elina Kaarlejärvi, Emilie Kissler, Timo Kumpula, Juul Limpens, Isla H. Myers-Smith, Signe Normand, Eric Post, Adrian V. Rocha, Niels Martin Schmidt, Anna Skarin, Eeva M. Soininen, Aleksandr Sokolov, Natalia Sokolova, James D. M. Speed, Lorna Street, Nikita Tananaev, Jean-Pierre Tremblay, Christine Urbanowicz, David A. Watts, Heike Zimmermann and Johan Olofsson

E. Lindén (https://orcid.org/0000-0002-4060-0110) ✉ (elin.linden@umu.se) and J. Olofsson, Dept of Ecology and Environmental Science, Umeå Univ., Umeå, Sweden. – M. te Beest, Copernicus Inst. for Sustainable Development, Utrecht Univ., Utrecht, the Netherlands and Centre for African Conservation Ecology, Nelson Mandela Univ., Port Elizabeth, South Africa. – I. Aubreu, Dept of Plant Biochemistry, Univ. of Göttingen, Göttingen, Germany. – IA and T. Moritz, Umeå Plant Science Centre, Dept of Forest Genetics and Plant Physiology, Swedish Univ. of Agricultural Sciences, Umeå, Sweden. – M. K. Sundqvist, Dept of Forest Ecology and Management, Swedish Univ. of Agricultural Science, Umeå, Sweden. – I. C. Barrio, Faculty of Environmental and Forest Sciences, Agricultural Univ. of Iceland, Reykjavik, Iceland. – J. Boike and H. Zimmermann, Alfred Wegener Inst., Hemholtz Centre for Polar and Marine Research, Potsdam, Germany. JB also at: Geography Dept, Humboldt-Univ. zu Berlin, Berlin, Germany. – J. P. Bryant, Inst. of Arctic Biology, Univ. of Alaska Fairbanks, Fairbanks, AK, USA. – K. A. Bråthen and E. M. Soininen, Dept of Arctic and Marine Biology, Faculty of Biosciences, Fisheries and Economics, UiT – The Arctic Univ. of Norway, Tromsø, Norway. – A. Buchwal, Inst. of Geoecology and Geoinformation, Adam Mickiewicz Univ., Poznan, Poland. – C. G. Bueno (https://orcid.org/0000-0002-7288-2271), Inst. of Ecology and Earth Sciences, Dept of Botany, Univ. of Tartu, Tartu, Estonia. – A. Currier, Jardin Botanique de Montréal, Institut de Recherche en Biologie Végétale, Université de Montréal, Montréal, QC, Canada. – D. D. Egelkraut, Univ. of Bergen, Dept of Biological Sciences, Bergen, Norway. – B. C. Forbes, Arctic Centre, Univ. of Lapland, Rovaniemi, Finland. – M. Hallinger, Inst. of Botany and Landscape Ecology, Univ. of Greifswald, Greifswald, Germany. – M. Heijmans, M. Holmgren and J. Limpens, Dept of Environmental Sciences, Wageningen Univ., Wageningen, the Netherlands. – L. Hermanutz and E. Kissler, Dept of Biology, Memorial Univ. of New Foundland, St. John's, NL, Canada. – D. S. Hik, Dept of Biological Sciences, Simon Fraser Univ., Burnaby, BC, Canada. – A. Hofgaard, Norwegian Inst. for Nature Research, Trondheim, Norway. – D. C. Huebner, Dept of Biology and Wildlife, Univ. of Alaska Fairbanks, Fairbanks, AK, USA. – T. T. Høye (https://orcid.org/0000-0001-5387-3284), Dept of Ecoscience and Arctic Research Centre, Aarhus Univ., Rønde, Denmark. – I. S. Jónsdóttir, Inst. of Life and Environmental Sciences, Univ. of Iceland, Reykjavík, Iceland. – E. Kaarlejärvi, Research Centre for Ecological Change, Organismal and Evolutionary Biology Research Programme, Univ. of Helsinki, Helsinki, Finland. – T. Kumpula, Dept of Geographical and Historical Studies, Univ. of Eastern Finland, Joensuu, Finland. JL also at: Plant Ecology and Nature Conservation, Wageningen, the Netherlands. – I. H. Myers-Smith and L. Street, School of GeoSciences, Univ. of Edinburgh, The King's Buildings, Edinburgh, UK. – S. Normand, Dept of Biology, Aarhus Univ., Aarhus C, Denmark. – E. Post (https://orcid.org/0000-0002-9471-5351), Dept of Wildlife, Fish and Conservation Biology, Univ. of California,

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Davis, CA, USA. – A. V. Rocha (https://orcid.org/0000-0002-4618-2407), Dept of Biological Sciences, Univ. of Notre Dame, Notre Dame, IN, USA. – Niels Martin Schmidt, Dept of Ecoscience, Aarhus Univ., Roskilde, Denmark and Arctic Research Centre, Aarhus Univ., Aarhus, Denmark. – A. Skarin (https://orcid.org/0000-0003-3221-1024), Dept of Animal Nutrition and Management, SLU, Uppsala, Sweden. – A. Sokolov and N. Sokolova (https:// orcid.org/0000-0003-4280-8350), Arctic Research Station of Inst. of Plant and Animal Ecology, Ural Branch, Russian Academy of Sciences, Labytnangi, Yamal-Nenets Autonomous District, Russia. – J. D. M. Speed (https://orcid.org/0000-0002-0633-5595), Dept of Natural History, NTNU Univ. Museum, Norwegian Univ. of Science and Technology, Trondheim, Norway. – N. Tananaev, Melnikov Permafrost Inst. SB RAS, Yakutsk, Russia. – J.-P. Tremblay, Dépt de Biologie, Faculté des Sciences et de Génie, Univ. Laval, Québec, Canada. – C. Urbanowicz, Dept of Biological Sciences, Dartmouth College, Hanover, NH, USA. – D. A. Watts (https://orcid.org/0000-0003-2054-7719), Alaska State Virology Laboratory, Division of Public Health, Alaska Dept of Health and Social Services, Fairbanks, Alaska, USA.

Spatial variation in plant chemical defence towards herbivores can help us understand variation in herbivore top–down control of shrubs in the Arctic and possibly also shrub responses to global warming. Less defended, non-resinous shrubs could be more influenced by herbivores than more defended, resinous shrubs. However, sparse field measurements limit our current understanding of how much of the circum-Arctic variation in defence compounds is explained by taxa or defence functional groups (resinous/non-resinous). We measured circum-Arctic chemical defence and leaf digestibility in resinous (*Betula glandulosa*, *B. nana* ssp. *exilis*) and non-resinous (*B. nana* ssp. *nana*, *B. pumila*) shrub birches to see how they vary among and within taxa and functional groups. Using liquid chromatography–mass spectrometry (LC–MS) metabolomic analyses and in vitro leaf digestibility via incubation in cattle rumen fluid, we analysed defence composition and leaf digestibility in 128 samples from 44 tundra locations.

We found biogeographical patterns in anti-herbivore defence where mean leaf triterpene concentrations and twig resin gland density were greater in resinous taxa and mean concentrations of condensing tannins were greater in non-resinous taxa. This indicates a biome-wide trade-off between triterpene- or tannin-dominated defences. However, we also found variations in chemical defence composition and resin gland density both within and among functional groups (resinous/non-resinous) and taxa, suggesting these categorisations only partly predict chemical herbivore defence. Complex tannins were the only defence compounds negatively related to in vitro digestibility, identifying this previously neglected tannin group as having a potential key role in birch anti-herbivore defence.

We conclude that circum-Arctic variation in birch anti-herbivore defence can be partly derived from biogeographical distributions of birch taxa, although our detailed mapping of plant defence provides more information on this variation and can be used for better predictions of herbivore effects on Arctic vegetation.

Introduction

In the absence of trees, the tallest vascular plants in the tundra are shrubs. They are currently increasing in abundance, height and distribution across the Arctic and global warming is a main driver of these changes (Myers-Smith et al. 2011, Bjorkman et al. 2018, Berner et al. 2020). This increase in shrubs may lead to a decline in plant diversity (Myers-Smith et al. 2011), decreased albedo (Sturm et al. 2005) and increased greenhouse gas emissions (Hartley et al. 2012) in tundra ecosystems. Arctic shrubification patterns are, however, highly variable across space and time and sometimes deviate from observed warming trends (Berner et al. 2020). A factor that can contribute to this variability is herbivory (Christie et al. 2015). Large herbivores such as caribou/ reindeer, sheep and muskoxen already slow down shrub expansion and growth in many Arctic ecosystems (Post and Pedersen 2008, Olofsson et al. 2009, Hofgaard et al. 2010, Bråthen et al. 2017), but not all (Tremblay et al. 2012). Insect herbivores can also cause severe defoliation and suppress woody plants during mass outbreak events (Jepsen et al. 2008, Prendin et al. 2019), but such events are rare in the tundra where even background insect herbivory is low (Barrio et al. 2017). How efficiently herbivores may limit Arctic shrub

expansion depends on shrub palatability and digestibility. This is partly linked to their growth rates and resource acquisition strategies, since more rapidly growing plants generally have nutrient-rich and thin (high specific leaf area) leaves (Wright et al. 2004). All shrubs also produce chemical anti-herbivore defence compounds (Kramer and Kozlowski 1979), with presumed deterrent and/or digestion-inhibiting effects on herbivores (Bryant et al. 1991, Christie et al. 2015).

Deciduous shrubs of the genus *Betula* (hereafter shrub birches or birches) are common in most vegetated areas in low Arctic tundra and are often subjects for ecological studies on shrub response to climate change. Their most important chemical defences against herbivores are suggested to be carbon-based triterpenes and condensed tannins (Bryant et al. 2014). At least some triterpene compounds are directly toxic to mammalian herbivores (McLean et al. 2009), while condensed tannins should primarily reduce plant nutritional quality by inhibiting protein digestion and are thus often categorised as digestive reducers (Julkunen-Tiitto et al. 1996). Other compounds present in birch (Bryant et al. 2014), and often considered in herbivore defence studies, are hydrolysable tannins that can cause oxidative stress in herbivores (Barbehenn et al. 2006), and flavonoids that can act as antifeedants but mainly protect plants from oxidative stress

and solar radiation (Kumar and Pandey 2013). Even within each compound group, there is a large diversity of different metabolites that can act as deterrents separately and/or have co-active effects (Gershenzon et al. 2012).

Defence compound composition can differ within species (Christie et al. 2015), as in the dwarf birch *Betula nana,* which includes subspecies that are either mainly terpene-defended (*B. nana* ssp. *exilis*) or mainly tannin-defended (*B. nana* ssp. *nana*). From a chemical plant defence perspective, these defence systems define two functional groups; resinous birches (e.g. *B. nana* ssp. *exilis*) and non-resinous birches (e.g. *B. nana* ssp. *nana*). Resinous birches are often considered to have a strong chemical defence, with twigs densely covered with resin glands that produce a resin rich in toxic dammarane triterpenes (McLean et al. 2009). These triterpenes show antifeeding effects on snowshoe hare and muskoxen when added to otherwise palatable food (Reichardt et al. 1984, White and Lawler 2002). Non-resinous birches, on the other hand, should lack the triterpene-rich resin and instead be predominantly defended by condensed tannins (Julkunen-Tiitto et al. 1996, Graglia et al. 2001). Condensed tannins are often presumed as weaker defence compounds compared to triterpenes, and non-resinous birches are therefore presumed to be more palatable to vertebrate herbivores than resinous birches (Bryant et al. 2014). A study directly comparing *B. nana* ssp. *nana* and *B. nana* ssp. *exilis* from two high- latitude sites suggests that non-resinous birches compensate for their lack of triterpenes by investing more carbon in greater production of tannins (Graglia et al. 2001). The generality of this trade-off across the Arctic is not known but, if present at larger spatial scales, this trade-off between triterpene and tannin defences is important for understanding how palatability of shrub birches varies across the Arctic and, by extension, to which degree their abundance and distribution may be limited by herbivory.

Bryant et al. (2014) proposed the hypothesis that herbivores may have a stronger effect on shrub birch abundance in areas with non-resinous birches (Fennoscandia, Iceland, Greenland and western Siberia) compared to areas dominated by resinous birches (most of the Canadian Arctic, Alaska and eastern Siberia), owing to their differences in chemical defence. Geographic variation in chemical defence could, therefore, be a major factor driving differences in top–down control of shrub birches by herbivores, and thus potentially explain variation in warming-driven shrubification patterns and greening across the Arctic (Bryant et al. 2014). Sparse field measurements currently limit our possibility to properly estimate how much of the circum-Arctic variation in anti-herbivore defence is actually explained by taxa or functional groups. Given that polyphenolic defence compounds can vary greatly at small spatial scales, between phenological phases (Torp et al. 2010) and over time (Salminen et al. 2002) within the same taxa, there is cause to believe that generalisations of chemical defence within taxa or functional groups might not be completely straightforward. More large-scale data measured across the climatic, geological and biotic gradients in Arctic tundra using comparable methods are needed to address this.

To test to what extent circum-Arctic variation in chemical defence composition in resinous and non-resinous birches can be explained by plant taxa and whether there is a trade-off between triterpene and tannin defence, we mapped the variation in chemical anti-herbivore defence in tundra shrub birches. We sampled resinous (*B. nana* ssp. *exilis*, *B. glandulosa*) and non-resinous (*B. nana* ssp. *nana*, *B. pumila*) shrub birches at 128 sampling sites within 44 locations across the circumpolar Arctic, and analysed concentrations of chemical defence compounds using liquid chromatography–mass spectrometry (LC–MS) metabolomics analysis. To determine an aspect of the importance of these defence compounds for vertebrate herbivores we also tested their effect on in vitro leaf digestibility. We hypothesise that: 1) Defence compound composition in tundra shrub birches shows more variation across the circumpolar Arctic than can be predicted based on taxa or functional group (resinous or non-resinous) alone. 2) There is a trade-off between triterpenes and tannins in shrub birches that can be detected at circum-Arctic scale, where non-resinous birches produce higher concentrations of tannins compared to triterpene-defended resinous birches that, instead, produce more triterpenes. 3) Tannins reduce leaf digestibility to a greater extent than triterpenes that deter herbivores through other mechanisms.

Material and methods

Study area and sampling

Our study covers circumpolar tundra vegetation in the Northern Hemisphere, at latitudes between 47.3°N and 74.5°N (Fig. 1), where sites (area of ~10 m radius) were selected based on shrub birch presence. During June–August 2014, we sampled resinous (*B. glandulosa*, *B. nana* ssp. *exilis*) and non-resinous dwarf birch (*B. nana* ssp. *nana*, *B. pumila*) at 128 sampling sites within 44 locations (Supporting information). Evenly distributed across each site, we randomly chose 10 individuals and sampled 50 random short-shoot leaves (leaf rosettes along the stem with leaves of the same age; n = 500 per site), and 10 random long-shoots (twigs and leaves representing the current annual growth; n = 100 per site) from each individual. All samples were air-dried in the field and further oven-dried at 60°C for 48 h upon arrival to the lab. We used the short-shoot leaves for chemical analyses to achieve as much phenological homogeneity as possible among samples, and long-shoot twigs for resin gland counts. Short-shoot leaf samples from the same site were pooled and ground with a ball mill prior to chemical analyses of defence compounds, total nitrogen (N), carbon (C) and in vitro digestibility.

Herbivores

The main large herbivores across our sites are reindeer or wild caribou (*Rangifer* sp.) with the exception of Zackenberg in eastern Greenland where muskoxen are the main herbivores and Audkuluheidi on Iceland where mainly sheep graze.
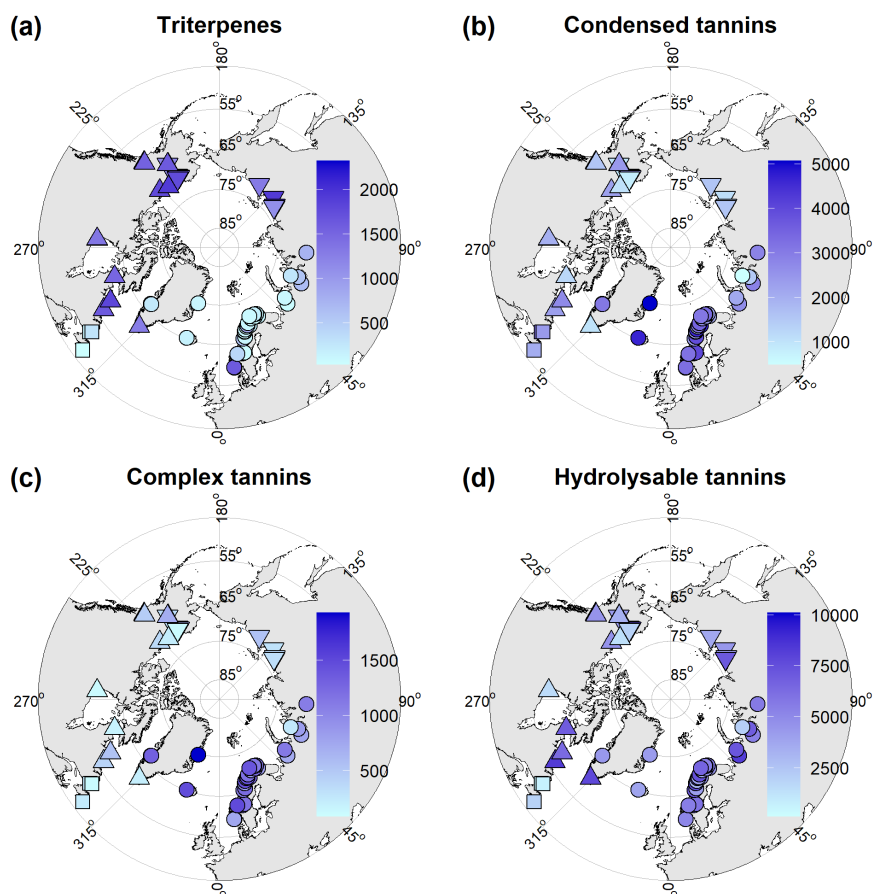
Figure 1. Maps showing relative concentrations (peak area/mg $\times$ 10^{-4} leaf tissue) of chemical compounds involved in the anti-browsing defence in tundra shrub birches. The maps cover high (dark blue) and low (light blue) concentrations of four compound classes; (a) triterpenes, (b) condensed tannins, (c) complex tannins and (d) hydrolysable tannins, in four tundra shrub birch taxa: *Betula glandulosa* (up-facing triangles), *B. nana* ssp. *exilis* (down-facing triangles), *B. nana* ssp. *nana* (circles) and *B. pumila* (squares).

Muskoxen also occur in other parts of Greenland, North America and Siberia, and moose might visit sites close to the treeline. Insect herbivores exist across the whole tundra, but none of our samples were taken in sites with active outbreaks.

Metabolite profiling

To profile chemical compounds connected to plant anti-browsing defence in tundra dwarf birch, we performed an untargeted metabolomics analysis by LC–MS at the Swedish Metabolomics Centre, Umeå Univ. We extracted 10 mg of ground leaf material, according to Gullberg et al. (2004), and analysed 2 µl of the extracts according to Abreu et al. (2020). Compounds were detected with an Agilent 6550 Q-TOF mass spectrometer with an electrospray ion source operating in negative ion mode. The MS files were processed by a targeted feature extraction using several in-house phenolic databases. Ionized mass from triterpenes was manually scanned across the MS spectra. When the same ion was present at a different retention time (e.g. due to in source fragmentation) it was annotated as 'derivative'. Metabolite identification was based on the diagnostic fragments produced during LC–MS analysis.

Defence compound groups

We identified approximately 100 metabolites (Supporting information), including several classes of defence compounds previously reported to contribute to plant anti-browsing defence such as triterpenes, condensed tannins, hydrolysable tannins, flavonoids and chlorogenic acid, as well as complex tannins. Complex tannins build up from a condensed tannin unit and a hydrolysable tannin unit, but are rarely considered or studied in ecological research. It is reasonable to assume complex tannins have similar protein-precipitating properties as condensed tannins, since they contain condensed tannin units. Flavonoids and chlorogenic acid (a precursor to phenolic compounds) are often considered of low importance as anti-feedants (Kumar and Pandey 2013), therefore they are mainly presented in the supplementary information of this study.

Resin gland density

To measure resin gland density, we defoliated 10 long-shoot twigs of each sample, photographed 15 mm twig segments starting 20 mm from the twig top and counted all visible resin glands. We measured the twig diameter at the beginning and

end of all segments to calculate the gland count area (approximated to one-half of the total segment bark area). We then calculated gland/mm² and used this as a standardised measure of resin gland density.

In vitro digestibility (IVOMD)

We measured general sample digestibility as in vitro organic matter digestibility (hereafter in vitro digestibility or IVOMD). Dry leaf material was mixed with rumen fluid from dairy cows and incubated at 38°C for 96 h, and the proportion of digested organic matter was then calculated, all according to Lindgren (1979). Although plant digestibility can be herbivore species-specific to some extent, overall differences in digestibility of food plants to different grazers and browsers can be estimated by in vitro organic matter digestibility with rumen fluid from dairy cows (Krizsan et al. 2018). The analyses were done at HUV Analysis Laboratory, Swedish Univ. of Agricultural Sciences, Uppsala.

Proxy of specific leaf area (SLA-P)

For all sampling sites, we weighed and scanned 10 random leaves from the pooled sample, and calculated the leaf area of dry leaves using the ROI (Region of Interest) manager tool in ImageJ software (Schneider et al. 2012). We used both long- and short-shoot leaves to standardise for fully developed leaves representing the whole birch individuals. Since our samples were dried upon arrival, we estimated a specific leaf area proxy (SLA-P) as (dry leaf area/dry mass) instead of SLA as (fresh leaf area/dry mass) (see Supporting information for results).

Nitrogen and carbon

Ground leaf samples were analysed for total nitrogen (N) and carbon (C) concentrations on an elemental analyser interfaced to a continuous flow isotope ratio mass spectrometer (IRMS, UC Davis Stable Isotope Facility, Univ. of California).

Climate

To relate the shrub taxa and their chemical defence composition to local climate conditions, we collected interpolated grid data with a resolution of 30 arc-seconds (~1 km) on annual precipitation, mean annual temperature, winter temperature (mean temperature of January) and summer temperature (mean temperature of July) from the WorldClim 1.4 database (Hijmans et al. 2005). Climate data for each sampling location were extracted using GIS software ArcMAP ver. 10.4.1 (ESRI 2011) (see Supporting information for relationships).

Statistical analyses

The chemical defence compounds were classified into six different groups of compounds; triterpenes, condensed tannins, hydrolysable tannins, complex tannins, flavonoids and chlorogenic acid (precursor to phenolic compounds). We included all compound groups for the statistical analyses to capture the overall diversity of chemical defence composition (hypothesis 1), but since triterpenes and tannins have the strongest contribution to plant defence we focus mainly on these in the main results (see Supporting information for additional information on flavonoids and chlorogenic acid). Similarities and differences in the chemical anti-herbivore defence composition across tundra shrub birches were explored by nonmetric multidimensional scaling analysis (NMDS, Minchin 1987) using the 'metaMDS' function of the 'vegan' package (Oksanen et al. 2019). In the NMDS we included all detected separate secondary metabolites from all six compound groups. To see if chemical defence composition relates to other defence-associated attributes, we fitted resin gland density, leaf digestibility and leaf N and C content to the NMDS using the 'envfit' function in the 'vegan' package. Climatic and temporal associations to chemical composition were also tested with the 'envfit' function (Supporting information). Additionally, we examined similarities and dissimilarities in chemical defence among the shrub birches using a cluster analysis with Euclidean distance and Ward's minimum variance clustering method ('hclust' function in the 'stats' package); data were mean centred and scaled by standard deviation per sample. We used one-way ANOVAs to test for intertaxon differences in chemical defence and other plant traits. The chemical defence was tested at the compound group level, so for this we summed all compounds within each of the six groups and tested for differences among species. When we assessed the relationship between triterpenes and tannins (hypothesis 2), we included both condensed tannins and complex tannins in the tannin group, given that complex tannins also consist of condensed tannin units. Hydrolysable tannins do not have any condensing effects and are therefore not included in these analyses. We tested the relationship between tannin and triterpene concentrations with a linear regression. We also used linear regressions to test the relationship between concentrations of the most important defence compounds (triterpenes, condensed, hydrolysable and complex tannins) and in vitro digestibility (hypothesis 3). For the significant relations, we examined taxon dependencies using additional two-way ANOVAs. All statistical analyses were performed using R software (ver. 4.0.3) (<www.r-project.org>).

Results

We found clear large-scale patterns in spatial distribution in three out of the four most important defence compound groups (triterpenes, condensed tannins, hydrolysable tannins, complex tannins; Fig. 1), though we also found circum-Arctic variation in defence compound concentration in tundra shrub birch leaves.

Triterpene concentrations were highest in eastern Siberia and most of North America, and very low in eastern Canada (island of Newfoundland), Greenland, Iceland, Scandinavia and western Siberia (Fig. 1a). We found the opposite pattern for condensed tannin and complex tannin
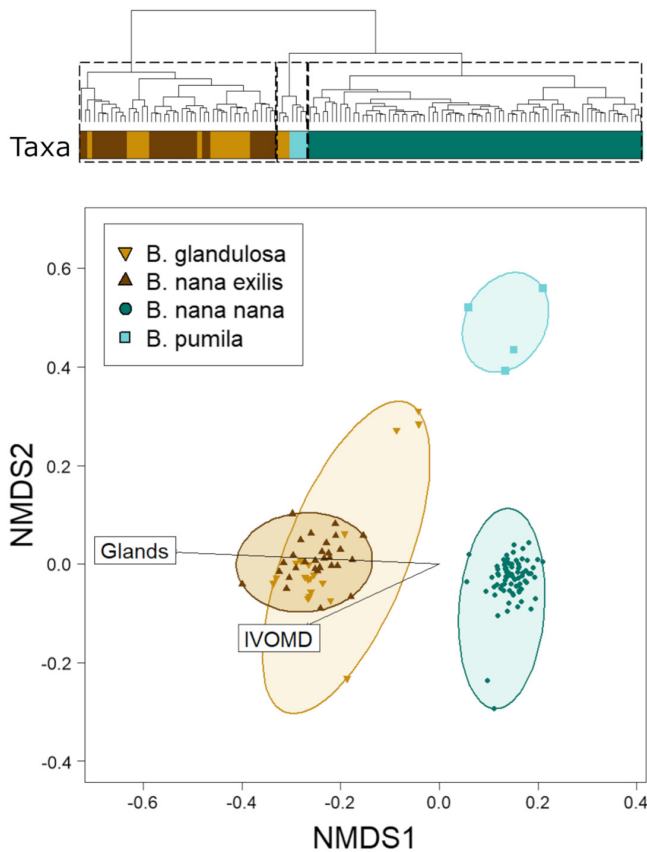
Figure 2. Non-metric dimensional scaling (NMDS) ordination and cluster diagram of chemical defence composition in resinous birches, *Betula glandulosa* (beige down-facing triangles) and *B. nana* ssp. *exilis* (brown up-facing triangles), and in non-resinous birches, *B. nana* ssp. *nana* (dark blue circles) and *B. pumila* (light blue squares). In the NMDS, ellipses represent the data range for each taxon, and black arrows show strength and direction of significant relationships between NMDS1 and resin gland density and in vitro digestibility. The coupled cluster diagram shows the three main clusters (in boxes) that form across the taxa based on chemical defence composition.

concentrations (Fig. 1b, c). Hydrolysable tannin concentrations varied less among biogeographical regions, except that the lowest concentrations of these compounds were found in Newfoundland, eastern Canada (Fig. 1d).

In the two-dimensional NMDS analysis (stress level = 0.09), higher scores on the first axis (NMDS1) were related to increasing tannin and decreasing terpene concentrations, while the second axis was mainly associated with a gradient in condensed to hydrolysable tannins, with higher condensed and lower hydrolysable tannin concentrations at high NMDS2 scores (Fig. 2). NMDS1 was negatively associated with higher resin gland density and invitro digestibility (IVOMD) (Fig. 2). The ranges of NMDS scores for each taxon showed that the non-resinous birches *B. pumila* and *B. nana* ssp. *nana* were clearly separated both from each other and from the resinous taxa (*B. nana* ssp. *exilis* and *B. glandulosa*), while the resinous birches had highly overlapping chemical composition. The IVOMD of birch leaves was

negatively related to complex tannins ($p = 0.003$, $R^2 = 0.07$), and positively related to triterpene ($p < 0.001$, $R^2 = 0.19$) and hydrolysable tannin ($p = 0.02$, $R^2 = 0.04$) concentrations (Fig. 3a, c, d). However, IVOMD was not related to condensed tannin concentrations (Fig. 3b). For the significant relations between IVOMD and separate compound group concentrations, we found intrataxon variation for triterpenes and complex tannins as well as both intra- and intertaxon variation for hydrolysable tannins. An additional cluster analysis revealed three main groupings across the taxa based on chemical defence composition, with *B. nana* ssp. *nana* as one group, *B. nana* ssp. *exilis* and most *B. glandulosa* as a second group and a third group with non-resinous *B. pumila* and a few resinous *B. glandulosa* samples (Fig. 2) (see Supporting information for more details on the chemical similarities and differences among samples).

The most apparent difference between the four birch taxa was the higher triterpene concentrations in the resinous birches *B. glandulosa* and *B. nana* ssp. *exilis*, compared to the non-resinous birches *B. nana* ssp. *nana* and *B. pumila* (Table 1, Fig. 4a). Additionally, *B. nana* ssp. *nana* had a higher condensed tannin concentration than the two resinous taxa *B. glandulosa* and *B. nana* ssp. *exilis*, while condensed tannin concentrations in *B. pumila* did not differ from the other taxa (Table 1, Fig. 4b). Hydrolysable tannin concentrations were higher in *B. nana* ssp. *exilis* than in *B. glandulosa*, while concentrations in *B. nana* ssp. *nana* were intermediate, and did not differ from the two resinous taxa (Table 1, Fig. 4d). In *B. pumila*, hydrolysable tannin concentrations were much lower than in any other taxa (Table 1, Fig. 4d). The complex tannin concentrations were higher in *B. nana* ssp. *nana* compared to the other three taxa, and lower in *B. pumila* compared to the two *B. nana* subspecies (Table 1, Fig. 4c). Flavonoid concentrations were highest in *B. nana* ssp. *nana* and lowest in *B. pumila* (Table 1; Supporting information), and there were no differences in chlorogenic acid concentrations between the four taxa (Table 1; Supporting information). In general, the variation within taxa was large for all compounds (even triterpenes), resulting in overlapping concentrations among taxa. Across all samples, triterpene concentration was negatively correlated to concentrations of condensed and complex tannins combined, indicating a compensatory production of these tannins in shrubs that lack triterpenes ($p < 0.001$, $R^2 = 0.29$) (Fig. 4e).

As expected, twig resin gland density was higher in the resinous taxa *B. glandulosa* and *B. nana* ssp. *exilis* than in the non-resinous taxa *B. nana* ssp. *nana* and *B. pumila*. This was clearly distinct, and gland density values do not overlap between the two functional groups (Table 1, Fig. 5a). IVOMD was highest in *B. glandulosa*, intermediate in *B. nana* ssp. *exilis* and *B. nana* ssp. *nana*, and lowest in *B. pumila*, but the differences between *B. pumila* and the two *B. nana* subspecies were not statistically significant (Table 1, Fig. 5b). N concentrations were similar among the four taxa, and even though they tended to be slightly higher in *B. nana* ssp. *exilis* than in *B. glandulosa* (Table 1, Supporting information), N concentrations did not explain any of the differences
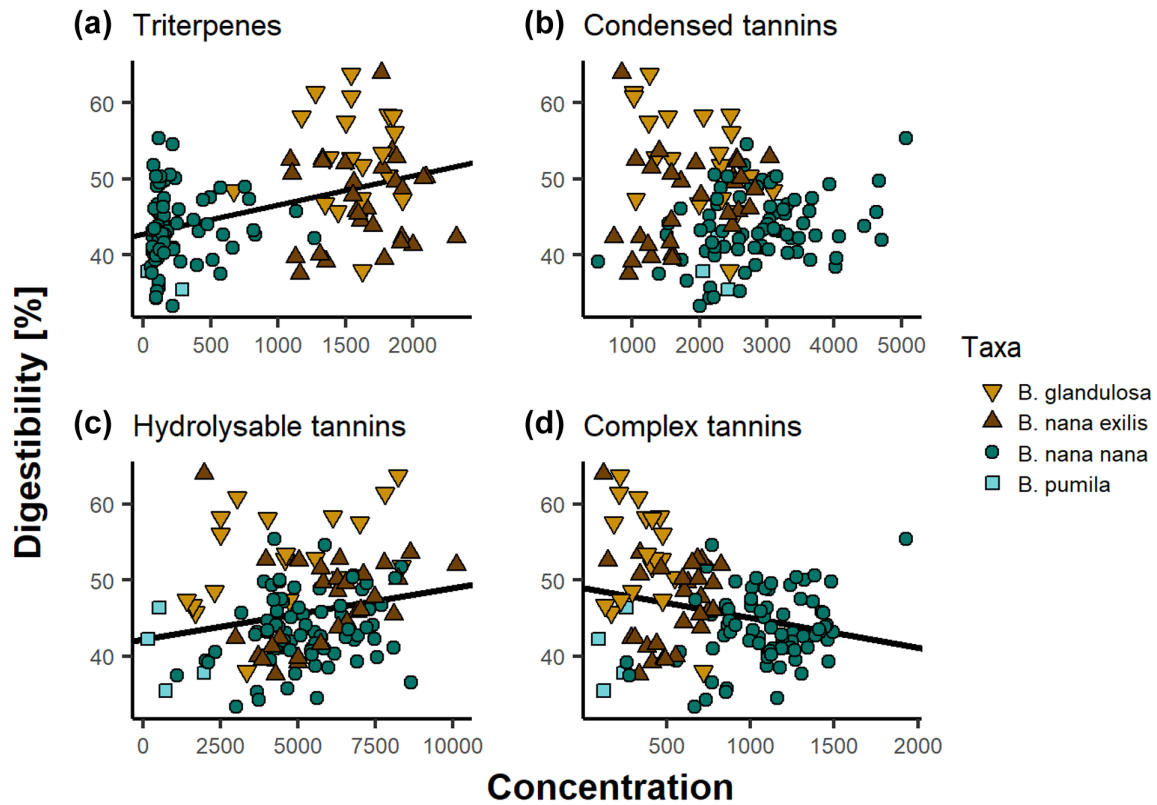
Figure 3. Linear relationships between digestibility (in vitro organic matter digestibility; (%) meltability of organic dry matter in cow rumen) and chemical defence compound concentration ((peak area/mg $\times 10^{-4}$) in (a) triterpenes, (b) condensed, (c) hydrolysable and (d) complex tannins) in four tundra shrub birch taxa: resinous *Betula glandulosa* (beige) and *B. nana* ssp. *exilis* (brown), and non-resinous *B. nana* ssp. *nana* (dark blue) and *B. pumila* (light blue). Significant linear relationships are drawn with bold lines.

in defence composition (Fig. 2). Also, there was no difference in mean C concentrations among the four taxa (Table 1, Supporting information).

Discussion

Our spatially and chemically detailed data on anti-herbivore defence in birch leaves reveal biogeographic patterns as well as considerable circum-Arctic variation in chemical defence both within and among different taxa of tundra shrub birches. As hypothesised, leaves from resinous birches (*B. nana* ssp. *exilis*, *B. glandulosa*) had higher triterpene concentrations than leaves from non-resinous taxa (*B. nana* ssp. *nana*, *B. pumila*). This finding was expected, since triterpenes are mostly produced in the resin glands (Bryant et al. 2014), and occurrence of resin glands is one of the characteristics separating resinous from non-resinous birches (<www.efloras.org/

Table 1. One-way ANOVAs testing intertaxon variation in chemical compounds and other attributes involved in plant anti-herbivore defence in four tundra shrub birch taxa (*Betula glandulosa*, *B. nana* ssp. *nana*, *B nana* ssp. *exilis*, *B. pumila*). Significant values in bold.

| Source of variation	Taxon variation		
	df	F	p
Triterpene	**3**	**200.1**	**< 0.001**
Condensed tannin	**3**	**15.75**	**< 0.001**
Hydrolysable tannin	**3**	**11.03**	**< 0.001**
Complex tannin	**3**	**73.14**	**< 0.001**
Flavonoids	**3**	**66.25**	**< 0.001**
Chlorogenic acid	3	1.05	0.373
Gland density	**3**	**232**	**< 0.001**
In vitro organic matter digestibility (IVOMD)	**3**	**3.18**	**0.027**
Carbon	3	0.76	0.517
Nitrogen	**3**	**2.97**	**0.034**
Residuals	124[a]		

[a] For all but 'Gland density' with residuals=123.
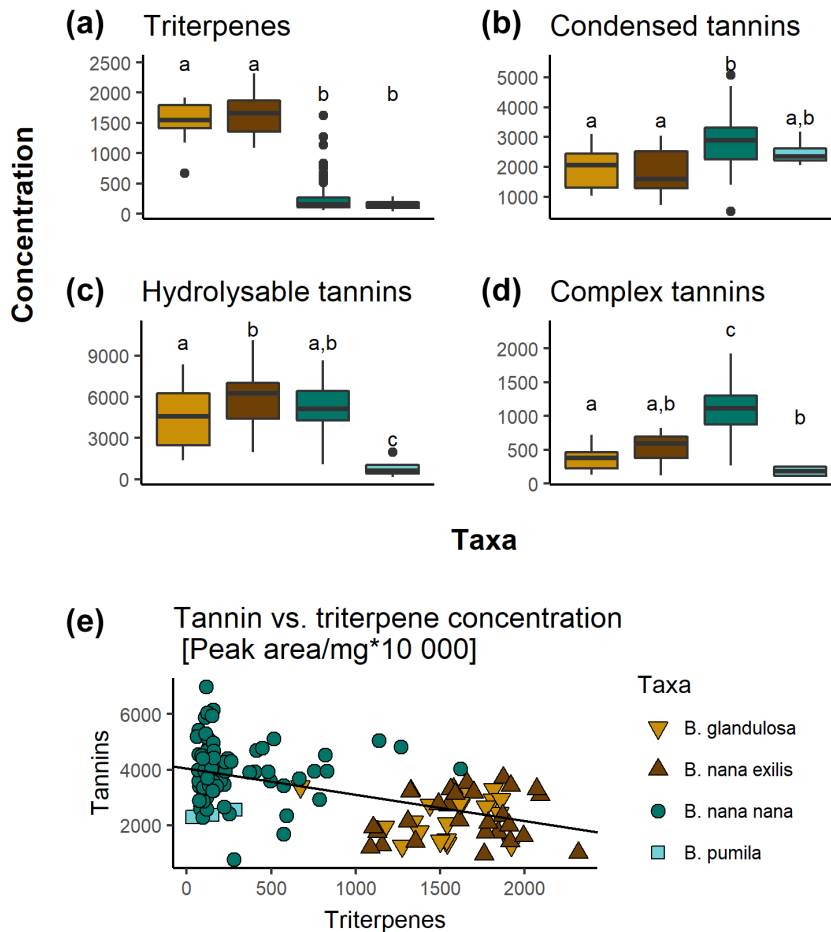
Figure 4. Boxplots showing (a) triterpene, (b) condensed tannin, (c) hydrolysable tannin and (d) complex tannin concentrations (peak area/mg × 10⁻⁴) in resinous *Betula glandulosa* (beige, n=19) and *B. nana* ssp. *exilis* (brown, n=33), and in non-resinous *B. nana* ssp. *nana* (dark blue, n=76) and *B. pumila* (light blue, n=4) tundra shrub birches. Boxplots show median (thick line), interquartile range (box), outer quartile (error bar) and outliers outside two interquartile ranges (dots). Significant differences in group means are annotated by lowercase letters. The scatterplot (e) shows the significant linear relationship between tannin (condensed+complex) and triterpene concentrations for all four birch taxa.

florataxon.aspx?flora_id=1&taxon_id=233500254www.eflo-ras.org/florataxon.aspx?flora_id=1&taxon_id=233500254>). Nevertheless, we also found support for hypothesis 1; variation in chemical defence composition across the Arctic was not fully explained by functional groups or taxa. In general,

we found the highest triterpene concentrations in the birch taxa with high resin gland density, although the differences in triterpene concentrations were not as pronounced as differences in resin glands on twigs. We also found a few non-resinous *B. nana* spp. *nana* samples that had relatively high
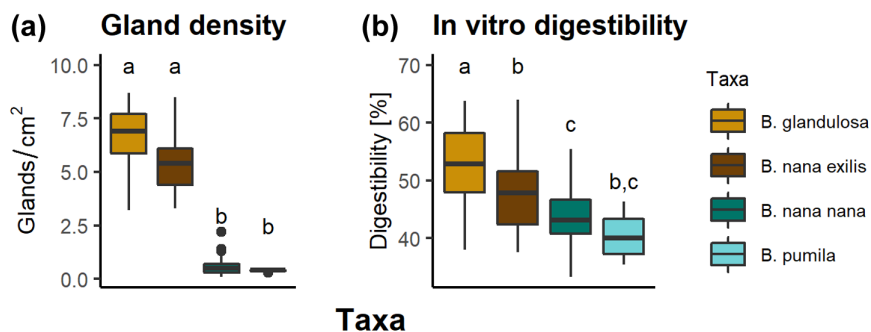

Figure 5. Boxplots showing twig resin gland density (a) and leaf in vitro digestibility (b) in resinous *Betula glandulosa* (beige, n=19) and *B. nana* ssp. *exilis* (brown, n=33), and in non-resinous *B. nana* ssp. *nana* (dark blue, n=76) and *B. pumila* (light blue, n=4) tundra shrub birches. The boxplots show median (thick line), interquartile (box), outer quartile (error bar) and outliers outside two interquartile ranges (dots). Significant differences in group means are annotated by lowercase letters.

leaf triterpene concentrations despite low gland density, suggesting that leaf triterpene concentrations are not always directly associated with the number of resin glands on twigs. Non-resinous birches can have a few resin-producing glands (Bryant et al. 2014), which in some cases in our study resulted in triterpene concentrations similar to some resinous birches. We also discovered that, despite being densely covered by glands, some resinous birches (*B. glandulosa*) may lack triterpenes almost entirely and instead have a chemical defence composition that is similar to the non-resinous and poorly gland-covered *B. pumila* (Supporting information). These findings indicate substantial variability in chemical defence by triterpenes that can be unrelated to resin gland density and result in overlaps in triterpene concentrations among resinous and non-resinous taxa. Thus, morphological characters such as resin glands are not alone sufficient to predict herbivore defence in leaves, and the geographical distribution of resinous/non-resinous birches cannot adequately predict to what extent shrub growth will be retarded by herbivores.

In agreement with our second hypothesis 2, we found evidence that non-resinous birch compensate for their lower concentrations of triterpenes by producing more condensing tannins through both higher condensed tannin production and higher complex tannin production. In this study, *B. nana* ssp. *nana* had the highest concentrations of both condensed and complex tannins of all birch taxa. Compared to its resinous counterpart, *B. nana* ssp. *exilis*, condensed tannin concentrations were more than 50% greater, and complex tannin concentrations more than twice as great, in *B. nana* ssp. *nana*. Our results confirm findings of condensed tannin concentrations being up to 50% higher in non-resinous *B. nana* ssp. *nana* from Abisko, Sweden, compared to resinous *B. nana* ssp. *exilis* from Toolik, Alaska (Graglia et al. 2001), and this study expands upon that finding by showing that this pattern may be widespread across the Arctic. We also found differences in tannin composition between the non-resinous birches. *B. pumila* are almost exclusively defended by condensed tannins, while *B. nana* ssp. *nana* is characterised by a more diverse defence system which also includes hydrolysable and complex tannins. Within the resinous birch taxon (*B. glandulosa* and *B. nana* ssp. *exilis*) we also found substantial variation in tannin composition, even though these two taxa, categorised as different species, cannot be separated on the basis of their defence composition. Also, despite the fact that sampling date did not influence plant defence concentrations in our study (Supporting information), temporal patterns are common for defence compound concentrations (Salminen et al. 2002). These findings emphasize that both non-resinous and resinous birch taxa can vary greatly in tannin composition across the Arctic, further indicating challenges for circum-Arctic generalisations of chemical defence composition.

Mapping the concentrations of plant defence compounds across the Arctic is essential for understanding how effects of trophic interactions and climate change on plant growth varies across the Arctic (Christie et al. 2015). Shrub birches are especially important in the Arctic because they are dominant in many parts of the tundra biome, respond quickly to warming and fertilisation (Chapin et al. 1995), yet can be affected by herbivory (Olofsson et al. 2009). Both triterpenes and tannins are complex compound groups, but they are still often suggested as deterrents of both vertebrate and insect herbivory (Barbehenn and Constabel 2011, Christie et al. 2015). The inhibitory functions of triterpenes on herbivores are diverse (Pichersky and Raguso 2018). They can, for example, be directly toxic by influencing the nervous system or by acting as hormone analogues in herbivores (Agrawal et al. 2012). Tannins are a diverse group of polyphenolics, and their effects on herbivores are even more variable (Salminen and Karonen 2011). Most tannins have the potential to bind and precipitate proteins (Marsch et al. 2020), but condensed tannins are the ones presumed to have the highest capacity to reduce protein digestion (Jayanegara and Palupi 2010). The effects of hydrolysable tannins are even less clear, but at least some reduce food digestibility by complex-binding sugar molecules (Goel et al. 2005), and some cause oxidative stress in herbivores (Barbehenn and Constabel 2011).

Our test of plant digestibility at least partly confirmed our hypothesis 3 that tannins reduce leaf digestibility while triterpenes do not since they deter herbivores through other mechanisms. Only one tannin group, complex tannins, showed a negative relationship to in vitro digestibility in this study. This highlights the possible importance of these compounds in chemical anti-herbivore defence. There are essentially no data on the effect of complex tannins on herbivores, but since they are made up of both hydrolysable and condensed tannins, it makes sense that they act as digestion reducers, as indicated by this study. Although complex tannins have not been considered as a separate group in earlier ecological studies, they have probably been included in many of the traditional quantification methods of condensed tannins, such as the acid–butanol (Porter et al. 1985) and vanillin (Price et al. 1978) assays, since their condensed tannin unit is likely targeted by these methods. The lack of a relationship between condensed tannins and digestibility in this study is thus not necessarily in conflict with previous studies that found a clear decreased leaf digestibility with higher condensed tannin concentrations (Gowda et al. 2019), since those authors used traditional quantification methods likely targeting complex tannins as well. Hydrolysable tannins and triterpenes were actually positively related to leaf in vitro digestibility in this study. It is not likely that these compounds stimulate digestion, and we interpret the relationships as spurious correlations caused by triterpenes and hydrolysable tannins being negatively correlated to complex and condensed tannins. The leaf digestibility in our in vitro inoculums in cow rumen ranged between 38% and 60%, which is consistent with earlier studies testing both in vitro and in vivo digestibility of *B. nana* ssp. *exilis* in caribou rumen (45 and 54%, respectively) (Kuropat 1984). Plant digestibility and palatability is, however, a product of complex plant syndromes (Agrawal and Fishbein 2006), resulting from strong associations among a wide range of different chemical and morphological attributes, and it is therefore hard to disentangle the causal effects of single components in composite samples.

The mapping of plant defence compounds presented in this study can be an important tool for predicting how shrub birches defend themselves against herbivory across the Arctic, and a prerequisite for future research. For example, the circum-Arctic range we find of the trade-off relation between triterpenes and condensed and complex tannins raises questions of the dose response of these compounds. Deriving effect equivalents or a shared defence currency among different compounds is needed to translate concentrations of different compounds to defence strength. Finding out the dose response of separate compound groups could also allow for estimates of relative costs for plants to produce certain types of defences and the cost for herbivores to cope with them, but data to do so are currently not available. Existing herbivore feeding trials do not fully resolve this question either, since deterrent effects of different compound groups are generally tested separately (Reichardt et al. 1984, White and Lawler 2002).

We are only aware of one study that compares relative palatability of non-resinous and resinous birches that showed that, in winter, free-ranging snowshoe hares (Alaska) and mountain hares (Finland) feed preferentially upon the less resinous birch when presented with twigs from both birch types (Bryant 1989). In this study, we reveal that tannin and terpene concentrations can vary substantially within both non-resinous and resinous birches, so multisite feeding trials would be needed to test the generality of these results. Resinous birches are not totally avoided by herbivores since herbivores can have strong suppressing effects also on the resinous birch *B. glandulosa* (Manseau et al. 1996, Crête and Doucet 1998, Andruko et al. 2020). Plant responses to herbivory are not only a product of plant chemistry, but they also depend on the ecological context in which plants grow. Herbivore density is an important aspect that is often controlled to a greater extent by climate, landscape structure, predators and human activities than by plant chemistry (Dahlgren et al. 2009, Skarin et al. 2020). Herbivore guild also matters, since the same compounds might have different effects on different herbivores (Barbehenn and Constabel 2011). The extent to which plants are exposed to herbivory is also related to the composition and diversity of neighbouring plants, since the quality of alternative resources can affect herbivore preference for certain plants (Barbosa et al. 2009) and herbivore attraction to certain areas (Palmer et al. 2003).

We conclude that although circum-Arctic variation in birch chemical defence can be partly explained by characterisation as taxon and defence type (resinous/non-resinous), there is additional variation in defence chemistry that these groupings do not explain. Detailed measurements of plant defence compounds, as in this study, can give a more accurate and complex description of the plant defence and thus provide a more powerful tool to predict future vegetation patterns. While chemical analyses alone will not give a full understanding of plant–herbivore interactions, our circum-Arctic mapping of defence compounds is a key contribution in understanding the strength of top–down control of shrub expansion in response to a warmer climate.

Author contributions

Elin Lindén: Conceptualization (equal); Data curation (lead); Formal analysis (lead); Investigation (lead); Methodology (equal); Project administration (lead); Visualization (lead); Writing – original draft (lead); Writing – review and editing (lead). **Mariska te Beest**: Conceptualization (equal); Investigation (supporting); Methodology (equal); Writing – original draft (supporting); Writing – review and editing (supporting). **Ilka Abreu**: Methodology (equal); Data curation (supporting); Investigation (supporting); Writing – original draft (supporting); Writing – review and editing (supporting). **Thomas Moritz**: Funding aquisition (supporting); Methodology (equal); Resources (supporting); Writing – review and editing (supporting). **Maja K. Sundqvist**: Conceptualization (equal); Investigation (supporting); Methodology (equal); Writing – original draft (supporting); Writing – review and editing (supporting). **Isabel C. Barrio**: Investigation (supporting); Writing – review and editing (supporting). **Julia Boike**: Investigation (supporting); Writing – review and editing (supporting). **John P. Bryant**: Investigation (supporting); Writing – review and editing (supporting). **Kari Anne Bråthen**: Investigation (supporting); Writing – review and editing (supporting). **Agata Buchwal**: Investigation (supporting); Writing – review and editing (supporting). **C. Guillermo Bueno**: Investigation (supporting); Writing – review and editing (supporting). **Alain Cuerrier**: Investigation (supporting); Writing – review and editing (supporting). **Dagmar D. Egelkraut**: Investigation (supporting); Writing – review and editing (supporting). **Bruce C. Forbes**: Investigation (supporting); Writing – review and editing (supporting). **Martin Hallinger**: Investigation (supporting); Writing – review and editing (supporting). **Monique Heijmans**: Investigation (supporting); Writing – review and editing (supporting). **Luise Hermanutz**: Investigation (supporting);

Writing – review and editing (supporting). **David S. Hik**: Investigation (supporting); Writing – review and editing (supporting). **Annika Hofgaard**: Investigation (supporting); Writing – review and editing (supporting). **Milena Holmgren**: Investigation (supporting); Writing – review and editing (supporting). **Diane C. Huebner**: Investigation (supporting); Writing – review and editing (supporting). **Toke T. Høye**: Investigation (supporting); Writing – review and editing (supporting). **Ingibjörg S. Jónsdóttir**: Investigation (supporting); Writing – review and editing (supporting). **Elina Kaarlejärvi**: Investigation (supporting); Writing – review and editing (supporting). **Emilie Kissler**: Investigation (supporting); Writing – review and editing (supporting). **Timo Kumpula**: Investigation (supporting); Writing – review and editing (supporting). **Juul Limpens**: Investigation (supporting); Writing – review and editing (supporting). **Isla H. Myers-Smith**: Investigation (supporting); Writing – review and editing (supporting). **Signe Normand**: Investigation (supporting); Writing – review and editing (supporting). **Eric Post**: Investigation (supporting); Writing – review and editing (supporting). **Adrian V. Rocha**: Investigation (supporting); Writing – review and editing (supporting). **Niels Martin Schmidt**: Investigation (supporting); Writing – review and editing (supporting). **Anna Skarin**: Investigation (supporting); Writing – review and editing (supporting). **Eeva M. Soininen**: Investigation (supporting); Writing – review and editing (supporting). **Aleksandr Sokolov**: Investigation (supporting); Writing – review and editing (supporting). **Natalya Sokolova**: Investigation (supporting); Writing – review and editing (supporting). **James D. M. Speed**: Investigation (supporting); Writing – review and editing (supporting). **Lorna Street**: Investigation (supporting); Writing – review and editing (supporting). **Nikita Tananaev**: Investigation (supporting); Writing – review and editing (supporting). **Jean-Pierre Tremblay**: Investigation (supporting); Writing – review and editing (supporting). **Christine Urbanowicz**: Investigation (supporting); Writing – review and editing (supporting). **David A. Watts**: Investigation (supporting); Writing – review and editing (supporting). **Heike Zimmermann**: Investigation (supporting); Writing – review and editing (supporting). **Johan Olofsson**: Conceptualization (equal); Formal analysis (supporting); Funding aquisition (lead); Investigation (supporting); Methodology (equal); Resources (supporting); Writing – original draft (supporting); Writing – review and editing (supporting)..

Data availability statement

Data are available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.9p8cz8whd> (Lindén et al. 2022).

Supporting information

The Supporting information associated with this article is available with the online version.

References

Abreu, I. N. et al. 2020. A metabolite roadmap of the wood-forming tissue in *Populus tremula*. – New Phytol. 228: 1559–1572.

Agrawal, A. A. and Fishbein, M. 2006. Plant defense syndromes. – Ecology 87: 132–149.

Agrawal, A. A. et al. 2012. Toxic cardenolides: chemical ecology and coevolution of specialized plant–herbivore interactions. – New Phytol. 194: 28–45.

Andruko, R. et al. 2020. Recent growth and expansion of birch shrubs across a low arctic landscape in continental Canada: are these responses more a consequence of the severely declining caribou herd than of climate warming? – Ecosystems 23: 1362–1379.

Barbehenn, R. V. and Constabel, C. P. 2011. Tannins in plant–herbivore interactions. – Phytochemistry 72: 1551–1565.

Barbehenn, R. V. et al. 2006. Tannin composition affects the oxidative activities of tree leaves. – J. Chem. Ecol. 32: 2235–2251.

Barbosa, P. et al. 2009. Associational resistance and associational susceptibility: having right or wrong neighbors. – Annu. Rev. Ecol. Evol. Syst. 40: 1–20.

Barrio, I. C. et al. 2017. Background invertebrate herbivory on dwarf birch (*Betula glandulosa-nana* complex) increases with temperature and precipitation across the tundra biome. – Polar Biol. 40: 2265–2278.

Berner, L. T. et al. 2020. Summer warming explains widespread but not uniform greening in the Arctic tundra biome. – Nat. Commun. 11: 4621.

Bjorkman, A. D. et al. 2018. Plant functional trait change across a warming tundra biome. – Nature 562: 57–62.

Bråthen, K. A. et al. 2017. Rangifer management controls a climate-sensitive tundra state transition. – Ecol. Appl. 27: 2416–2427.

Bryant, J. P. et al. 1989. Biogeographic evidence for the evolution of chemical defense by boreal birch and willow against mammalian browsing. – Am. Nat. 134: 20–34.

Bryant, J. P. et al. 1991. Interactions between woody plants and browsing mammals mediated by secondary metabolites. – Annu. Rev. Ecol. Syst. 22: 431–446.

Bryant, J. P. et al. 2014. Can antibrowsing defense regulate the spread of woody vegetation in arctic tundra? – Ecography 37: 204–211.

Chapin, F. S. et al. 1995. Responses of Arctic tundra to experimental and observed changes in climate. – Ecology 76: 694–711.

Christie, K. S. et al. 2015. The role of vertebrate herbivores in regulating shrub expansion in the arctic: a synthesis. – BioScience 65: 1123–1133.

Crête, M. and Doucet, G. J. 1998. Persistent suppression in dwarf birch after release from heavy summer browsing by caribou. – Arctic Alpine Res. 30: 126–132.

Dahlgren, J. et al. 2009. Plant defences to no avail? Responses of plants of varying edibility to food web manipulations in a low arctic scrubland. – Evol. Ecol. Res. 11: 1189–1203.

ESRI 2011. ArcGIS Desktop: release 10. – Environmental Systems Research Inst.

Gershenzon, J. et al. 2012. Mixtures of plant secondary metabolites: metabolic origins and ecological benefits. – In: Iason, G. R. et al. (eds), The ecology of plant secondary metabolites: from genes to global processes. Ecological reviews. Cambridge Univ. Press, pp. 56–77.

Goel, G. et al. 2005. Interaction of gut microflora with tannins in feeds. – Naturwissenschaften 92: 497–503.

Gowda, J. H. et al. 2019. Seasonal variation in the nutritional value of woody plants along a natural gradient in Eastern Africa. – Afr. J. Ecol. 57: 226–237.

Graglia, E. et al. 2001. Environmental control and intersite variations of phenolics in *Betula nana* in tundra ecosystems. – New Phytol. 151: 227–236.

Gullberg, J. et al. 2004. Design of experiments: an efficient strategy to identify factors influencing extraction and derivatization of Arabidopsis thaliana samples in metabolomic studies with gas chromatography/mass spectrometry. – Anal. Biochem. 331: 283–295.

Hartley, I. P. et al. 2012. A potential loss of carbon associated with greater plant growth in the European Arctic. – Nat. Clim. Change 2: 875–879.

Hijmans, R. J. et al. 2005. Very high resolution interpolated climate surfaces for global land areas. – Int. J. Climatol. 25: 1965–1978.

Hofgaard, A. et al. 2010. Comparing warming and grazing effects on birch growth in an alpine environment – a 10-year experiment. – Plant Ecol. Divers. 3: 19–27.

Jayanegara, A. and Palupi, E. 2010. Condensed tannin effects on nitrogen digestion in ruminants: a meta-analysis from in vitro and in vivo studies. – Media Peternakan 33: 176–181.

Jepsen, J. U. et al. 2008. Climate change and outbreaks of the geometrids *Operophtera brumata* and *Epirrita autumnata* in subarctic birch forest: evidence of a recent outbreak range expansion. – J. Anim. Ecol. 77: 257–264.

Julkunen-Tiitto, R. et al. 1996. Chemical diversity of several Betulaceae species: comparison of phenolics and terpenoids in northern birch stems. – Trees 11: 16–22.

Kramer, P. J. and Kozlowski, T. T. 1979. Physiology of woody plants. – Academic Press.

Krizsan, S. J. et al. 2018. An in vitro evaluation of browser and grazer fermentation efficiency and microbiota using European moose spring and summer foods. – Ecol. Evol. 8: 4183–4196.

Kumar, S. and Pandey, A. K. 2013. Chemistry and biological activities of flavonoids: an overview. – Sci. World J. 2013: 162750.

Kuropat, P. J. 1984. Foraging behavior of caribou on a calving ground in northwestern Alaska. – MSc thesis, Univ. of Alaska Fairbanks, AK, USA.

Lindén, E. et al. 2022. Data from: Circum-Arctic distribution of chemical anti-herbivore compounds in Arctic shrubs. – Dryad Digital Repository, <https://doi.org/10.5061/dryad.9p8cz8whd>.

Lindgren, E. 1979. Vallfodrets näringsvärde bestämt in vivo och med olika laboratoriemetoder (The nutritional value of roughages determined in vivo and by laboratory methods). – Swedish Univ. of Agricultural Science, Dept of Animal Husbandry, Report 45.

Manseau, M. et al. 1996. Effects of summer grazing by caribou on composition and productivity of vegetation: community and landscape level. – J. Ecol. 84: 503–513.

Marsch, K. J. et al. 2020. New approaches to tannin analysis of leaves can be used to explain in vitro biological activities associated with herbivore defence. – New Phytol. 225: 488–498.

McLean, S. et al. 2009. Papyriferic acid, an antifeedant triterpene from birch trees, inhibits succinate dehydrogenase from liver mitochondria. – J. Chem. Ecol. 35: 1252–1261.

Minchin, P. R. 1987. An evaluation of the relative robustness of techniques for ecological ordination. – Vegetatio 69: 89–107.

Myers-Smith, I. H. et al. 2011. Shrub expansion in tundra ecosystems: dynamics, impacts and research priorities. – Environ. Res. Lett. 6: 045509.

Oksanen, J. et al. 2019. vegan: community ecology package. – R package ver. 2.5-5, <https://CRAN.R-project.org/package=vegan>.

Olofsson, J. et al. 2009. Herbivores inhibit climate-driven shrub expansion on the tundra. – Global Change Biol. 15: 2681–2693.

Palmer, S. C. F. et al. 2003. The perils of having tasty neighbours: grazing impacts of large herbivores at vegetation boundaries. – Ecology 84: 2877–2890.

Pichersky, E. and Raguso, R. A. 2018. Why do plants produce so many terpenoid compounds? – New Phytol. 220: 692–702.

Porter, L. J. et al. 1985. The conversion of procyanidins and prodelphinidins to cyanidin and delphinidin. – Phytochemistry 25: 223–230.

Post, E. and Pedersen, C. 2008. Opposing plant community responses to warming with and without herbivores. – Proc. Natl Acad. Sci. USA 105: 12353–12358.

Prendin, A. L. et al. 2019. Immediate and carry-over effects of insect outbreaks on vegetation growth in West Greenland assessed from cells to satellite. – J. Biogeogr. 47: 87–100.

Price, M. L. et al. 1978. A critical evaluation of the vanillin reaction as an assay for tannin in sorghum grain. – J. Agric. Food Chem. 26: 1214–1218.

Reichardt, P. B. et al. 1984. Defense of winter-dormant Alaska paper birch against snowshoe hares. – Oecologia 65: 58–69.

Salminen, J.-P. and Karonen, M. 2011. Chemical ecology of tannins and other phenolics: we need a change in approach. – Funct. Ecol. 25: 325–338.

Salminen, J.-P. et al. 2002. Distribution of hydrolysable tannins in the foliage of Finnish birch species. – Z. Naturforsch. C 57: 248–256.

Schneider, C. A. et al. 2012. NIH Image to ImageJ: 25 years of image analysis. – Nat. Methods 9: 671–675.

Skarin, A. et al. 2020. Reindeer use of low Arctic tundra correlates with landscape structure. – Environ. Res. Lett. 15: 115012.

Sturm, M. 2005. Changing snow and shrub conditions affect albedo with global implications. – J. Geophys. Res. 110: G1.

Torp, M. et al. 2010. The effect of snow on plant chemistry and invertebrate herbivory: experimental manipulations along a natural snow gradient. – Ecosystems 13: 741–751.

Tremblay, B. et al. 2012. Recent expansion of erect shrubs in the low Arctic: evidence from Eastern Nunavik. – Environ. Res. Lett. 7: 035501.

White, R. G. and Lawler, J. P. 2002. Can methane suppression during digestion of woody and leafy browse compensate for energy costs of detoxification of plant secondary compounds? A test with muskoxen fed willows and birch. – Comp. Biochem. Physiol. A Mol. Integr. Physiol. 133: 849–859.

Wright, I. J. et al. 2004. The worldwide leaf economics spectrum. – Nature 428: 821–827.